however, severe distortion of the helix; 12,19 consistent with our data (vide supra), DNA melting curves<sup>18</sup> show the relative lability of the duplex structure.

Gel sequence analysis<sup>20</sup> of the platinated oligomer (Table II; Chart I) revealed a threefold increase in BLM-mediated damage at T<sub>6</sub>, which became the primary site of damage; a significant increase in cleavage at T<sub>16</sub> was also noted. The emergence of TT as a preferred Fe(II)·BLM cleavage site is unprecedented and provides an opportunity to identify those features in DNA that render the duplex susceptible to Fe·BLM-mediated modification.

The observed changes in sequence specificity are entirely consistent with those reported earlier with the use of cis-DDP and much longer DNA fragments.<sup>13</sup> However, this study extends the earlier findings in important ways, including better definition of the spatial relationships between the Pt center and the BLM cleavage sites that leads to altered cleavage patterns, the demonstration that conformational alteration in DNA can lead to novel BLM-mediated DNA cleavage patterns, and the finding that platination does not alter the chemistry of DNA cleavage.

Acknowledgment. We thank Dr. Robert Hertzberg, SK&F Laboratories, for helpful discussions and for assistance with characterization of the oligonucleotides. This study was supported by PHS Grant CA38544, awarded by the National Cancer Institute, D.H.H.S., and by an American Cancer Society Scholar Grant to B.G.

Registry No. BLM A2, 11116-31-7; BLM, 11056-06-7; cis-DEP, 14096-51-6; 5'-d(CGCT<sub>3</sub>A<sub>3</sub>CGC), 113403-23-9; Pt[d(CGCT<sub>3</sub>A<sub>3</sub>G<sub>2</sub>)]-d- $(C_2T_3A_3GCG)$ , 113403-26-2.

## Additions and Corrections

Determination of the Absolute Configurations of the Isomers of Triamminecobalt(III) Adenosine Triphosphate [J. Am. Chem. Soc. 1986, 108, 4167-4171]. DAVID C. SPECKHARD, VINCENT L. PECORARO, WILSON B. KNIGHT, and W. W. CLELAND\*

The assignments at the  $\alpha$ -phosphate were inadvertently switched, so that endo isomers are really exo and vice versa. The  $\beta$ -phosphate assignments are correct. Thus isomers 1–4 (in order of elution from cycloheptaamylose) correspond to  $\Lambda$ -exo,  $\Delta$ -exo,  $\Delta$ -endo, and  $\Lambda$ -endo, respectively.

Nitroethylene Yields N.N-Dihydroxyiminium-Methylium Dication in Trifluoromethanesulfonic Acid. Dications Stabilized by Y **Delocalization** [J. Am. Chem. Soc. 1987, 109, 7036]. Томоніко OHWADA, AKIKO ITAI, TOSHIHARU OHTA, and KOICHI SHUDO\* Page 7038, reference 12 (p7038) should read as follows:

In the case of planar conformations of N,N-dihydroxyiminium-methylium dication total energies obtained by MP2/ 6-31G\*//4-31G and relative energies (in parentheses) are as follows: 17, -282.6202686 au (0.00 kcal/mol); 18, -282.6122271 au (5.05 kcal/mol); 19, -282.603404 au (10.58 kcal/mol). (The other descriptions are deleted. This correction, however, does not make any revision in the discussion and conclusion described in the text.)

<sup>(19) (</sup>a) Macquet, J.-P.; Butour, J.-L. Biochemie 1978, 60, 901. (b) Cohen, G. L.; Bauer, W. R.; Barton, J. K.; Lippard, S. J. Science (Washington, D. C.) 1979, 203, 1014. (c) Cohen, G. L.; Ledner, J. A.; Bauer, W. R.; Ushay, H. M; Carvana, C.; Lippard, S. J. J. Am. Chem. Soc. 1980, 102, 2487. (d) Royer-Pokora, B.; Gordon, L. K.; Haseltine, W. A. Nucleic Acids. Res. 1981, 9, 4595. (e) Ushay, H. M.; Tullius, T. D.; Lippard, S. J. Biochemistry 1981, 20, 3744. (f) Scovell, W. M.; Caponi, V. J. Biochem. Biophys. Res. Commun. 1982, 107, 1138.

(20) Maxam, A. M; Gilbert, W. Methods Enzymol. 1980, 65, 499.